Formation of zygote in embryo sac

l'm not robot!









MORULA VERSUS DIASTILIA	
Morula is a solid ball of cells resulting from division of a fertilized ovum, and from which a blastula is formed	Blastula is a hollow sphere of cells surrounding the blastocoele produced during the development of an embryo
Forms 4-5 days after fertilization	Forms 5-10 days after fertilization
A solid cell mass	A hollow structure
Consists of a ball of small, spherical cells formed by the rapid cleavage of the zygote	Consists of a spherical cell layer of blastomeres and a fluid-filled cavity called blastocoel
Consists of more than a hundred of cells	Consists of hundred and twenty-three cells
Develops into the blastula in a process called blastulation	Develops into the gastrula in a process called gastrulation Visit www.pediaa.com

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Copyright © González-Gutiérrez et al.; licensee Springer. 2014 This article is published under license to BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited. Agave tequilana is an angiosperm species that belongs to the family Asparagaceae (formerly Agavaceae). Even though there is information regarding to some aspects related to the megagametogenesis of A. tequilana, this is the first report describing the complete process of megasporogenesis, megagametogenesis, the early embryo and characterize all the above processes and the distinctive morphological changes of the micropylar and chalazal extremes after fertilization in this species. The agave plant material for the present study was collected from commercial plantations in the state of Jalisco, Mexico. Ovules and immature seeds, previously fixed in FAA and kept in ethanol 70%, were stained based on a tissue clarification technique by using a Mayer's-Hematoxylin solution. The tissue clarification technique was successfully used for the characterization of the megasporogenesis, megagametogenesis, megagametogenesis, megagametogenesis, megagametogenesis, metare embryo sac formation. Also, the time-lapse of the developmental processes studied was recorded. The online version of this article (doi:10.1186/2193-1801-3-575) contains supplementary material, which is available to authorized users. Keywords: Agavaceae, Chalazal haustorium, Helobial endosperm, Hypostase, Megagametogenesis, Megagametogenesis, Polygonum-typeOne of the main characteristics of Angiosperms is that they possess seeds enclosed inside a fruit derived from the ovary of flowers (Li and Ma 2002). Another outstanding characteristic of angiosperms is that they present alternation of generations in their life cycle (as in many other plants), that is divided in two phases: one dominant diploid phase, which is called sporophytic, and one haploid phase known as gametophytic (Haig 1990; Rodríguez-Garay et al. 2008; Ma and Sundaresan 2010). The main function of the gametophyte phase is the production of haploid gametes whether they are male or female (Reiser and Fischer 1993; Yadegari and Drews 2004; Fan et al. 2008). The female gametophyte, also named megagametophyte or embryo sac, is developed within the carpel, which consists of three elements: the stigma, the style and the ovary, which can contain one or several ovules (Gutiérrez-Mora et al. 2012). In each ovule meiosis of the megaspore mother cell produces four haploid cells called megaspores. In the monosporic pattern, three of these megaspores degenerate while the closest cell to the chalazal region remains viable and gives rise to a single functional megaspore passes through one or more mitotic divisions without cytokinesis forming a multinucleate coenocyte. Latter, cell walls are formed around the nuclei resulting in a mature embryo sac (Rabiger and Drews 2013). The embryo sacs may present a diversity of developmental pathways, however, the most common is the monosporic Polygonum-type, in which the functional megaspore passes through three mitotic divisions producing a seven celled embryo sac (Chasan and Walbot 1993; Li and Ma 2002; Maheshwari 1937) consisting of three antipodal cells, one central cell formed by two polar nuclei, two synergid cells, and the egg cell (Dresselhaus 2006; Kägi and Groß-Hardt 2007; Yang et al. 2010). Some studies have characterized the female gametophyte of different species belonging to the Asparagaceae family formerly Agavaceae (APG III 2009), where the majority of them have been described as Monosporic Polygonum-type. Among these species, Yucca rupicola (Watkins 1937); Y. aloifolia (Wolf 1940); Y. filamentosa (Reed 1903); Agave lechuguilla (Grove 1941), A. virginica (Regen 1941), A. virginica (Regen 1941), A. virginica (Regen 1940); Y. filamentosa (Reed 1903); Agave lechuguilla (Grove 1948) and Comospermum yedoense (Rudall 1999) are found. However, Piven et al. (2001) reported the embryo sac development of Agave fourcroydes and A. angustifolia as bisporic Allium-type. Only one study could be found that the megagametophyte is originated from the megaspore that is located closest to the chalazal region, forming an embryo sac of seven cells, nor the early embryogenesis. Even though there is information regarding to some aspects related to the megagametogenesis of A. tequilana, there is no published information describing the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in this species and the distinctive morphological changes of the embryo and endosperm development in the distinctive morphological changes of the embryo and endosperm development in the distinctive morphological changes of the embryo and endosperm development in the distinctive morphological changes of the embryo and endosperm development in the distinctive morphological changes of the embryo and endosperm development in the distinctive morphological changes of the embryo and endosperm development in the distinctive morphological changes of the embryo and endosperm development in the distinctive morphological changes of the embryo and endosperm development in the distinctive morphological changes of the embryo and endosperm development in the distinctive development in the distinctive development in megagametogenesis, the mature embryo sac formation and the early embryo development in Agave tequilana Weber which is the raw material for the production of Tequila in Mexico, in order to get basic knowledge useful for plant systematics and evolution studies and plant breeding programs, which may include in vitro fertilization and the production of haploid plants among others. A total of 5,000 ovules were taken from floral buds at diverse stages, receptive flowers and immature fruits were analyzed. Different developmental stages of the collected ovules were studied from megasporogenesis to the first division of the embryo. The plant material used in this study consisted of the collected ovules were studied from megasporogenesis to the first division of the embryo. inflorescences collected from May to June in the years 2010-2013 (Figure 1). It was difficult to know the specific timeframe of each developmental stage of the embryo sac, however, it could be observed that it takes about 15 days for the floral buds since their appearance to reach the maturity of the embryo sac. Plant material of Agave tequilana Weber for the present study. a) Commercial plantation in the state of Jalisco, México. b) Hermafrodite agave flower. Bar = 1 cm. c) Agave ovary components. Bar = 50 µm. a = anthers, t = tepals, io = inferous ovary, s = style, o = ovules, lo = locules, p = placentae, ch = chalaza, m = micropyle, ii = interior integument, oi = outer integument, n = nucellar tissue, es = embryo sac. The process of megasporogenesis was observed using ovules in differentiation from the nucellar tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. The first results integrate tissue, es = embryo sac. 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The first results integrate tissue a dyad and finally in a linear tetrad of haploid cells with a chalaza-micropylar orientation (Figure 2b, c), the average size of the tetrad is formed, the three megaspores closest to the micropylar pole degenerate (Figure 2d) and their remains could be observed as highly stained spots which were frequently observed in the two-nucleated embryo sacs. Only the megaspore cell closest to the chalazal pole remains viable, becoming the functional megaspore (FM), and its size is bigger than the rest of the cells in the tetrad with dimensions of about 33 µm long and 23 µm wide. Megasporegenesis of Agave tequilana Weber. a) Diploid megaspore mother cell. b) Diad after the first meiotic division formed by four haploid cells. c) Tetrad after the second meiotic division formed by four haploid cells. c) Tetrad after the second meiotic division formed by four haploid cells. c) Tetrad after the second meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the second meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the second meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four haploid cells. c) Tetrad after the first meiotic division formed by four happendities after the first meiotic division formed by of the future embryo sac. ch = chalaza, m = micropyle, mmc = megaspore mother cell, d = dyad, t = tetrad, chm = chalazal megaspores. Bars = 10 µm. The process of megagametogenesis starts to be visible just above the FM in the nucellar tissue; this structure was detected as a well defined and intensely stained group of cells, which have a poor cytoplasmic content and thickened cell walls (Figure 3a) (also see Additional file 2: Figure S1). The first mitotic division of the functional megaspore produces two nuclei, one located at the chalazal pole and the other one at the micropylar pole, both separated by a large vacuole in the center of the embryo sac without cytokinesis. At this stage, the mean size of the embryo sac is about 50 µm wide (Figure 3b). Later, these two nuclei divided again forming four nuclei, two located at the chalazal end and the other two at the micropylar end of the embryo sac (Figure 3c). At the same time, the embryo sac continues expanding its size to about 60 µm long and 42 µm wide, and in both stages of embryo sac (Figure 3d). These three mitotic divisions occur in a synchronized manner at both extremes of the embryo sac. Megagametogenesis of Agave tequilana Weber. a) Functional megaspore. Bar = 20 µm. b) Two nuclei resulting after a first mitotic division. Bar = 20 µm. d) Eight-nuclei state forming a multinucleate coenocyte after the third and last mitotic division. Bar = 20 µm. ch = chalaza, hi = hypostase, m = micropylar polar nucleus, cni = chalazal nuclei, mni = micropylar nuclei. Arrowheads = eight final nuclei. The study of mature embryo sacs was carried out in ovules from flowers with mature and receptive stigma. At the mature stage, the size of the embryo sacs is about 247 µm long and 106 µm wide and they are wider at the chalazal pole than at the micropylar end, having a bulbous form with a small and narrow haustorial tube at the chalazal end which connects to the hypostase (Figure 4a). At this stage, the embryo sac is already cellularized and consists of seven cells: three antipodal cells could be observed); thee antipodal cells could be observed); the central cell formed by two polar nuclei was observed just below the antipodals and they were beside each other, being very similar in shape and size (Figure 4b) (also see Additional file 2: Figures S2 and S4). Finally, the egg apparatus composed of one egg cell and two synergids was observed at the micropylar end (Figure 4a, b and c). Mature embryo sac or megagametophyte of Agave tequilana Weber. a) Whole mature embryo sac showing the contents of both the chalazal and the micropylar poles. b) Polar nuclei at the chalazal end before karyogamy. c) Central cell nucleus after karyogamy took place. d) Egg apparatus composed by two synergid cells and the egg cell. ch = chalaza, ea = egg apparatus, m = micropyle, da = degenerating antipodal cell, ccn = central cell nucleus, ccv = central cell vacuole, pni = polar nuclei, s = synergid cells. Bars = 20 µm. Both synergids were found located at the micropylar end next to each other and generating antipodal cells. Bars = 20 µm. Both synergids were found located at the micropylar end next to each other and generating antipodal cells. Bars = 20 µm. Both synergids were found located at the micropylar end next to each other and generating antipodal cells. the egg cell was usually in a different focal plane and positioned between the two synergids. These three cells composing the ovular apparatus of Agave tequilana had their walls in contact with the micropylar edge of the female gametophyte (Figure 4d). The synergids are very similar to each other, and each cell possesses a vacuole polarized towards of Agave tequilana had their walls in contact with the micropylar edge of the female gametophyte (Figure 4d). the chalazal end and the nucleus polarized towards the micropylar end (see Aditional file 2: Figure S3). The egg cells showed diverse morphologies probably due to different developmental stages at the moment of observation, however, in general, they had a highly dense nucleus located towards the chalazal extreme and a large vacuole occupying almost the whole space of the cell located towards the micropylar end of the sac with a size of about 28 µm long and 23 µm wide (Figure 5). Different egg cell showing a highly condensed nucleus located at the chalazal end of the embryo sac. b) and c) Egg cells of an irregular shape with nucleus located at the chalazal end of the embryo sac. m = micropyle, ec = egg cell, ecn = egg cell nucleus. Bars = 20 µm. At the second day after pollination, the analyzed ovules did not show any change in size or morphology; however, the fusion of the two polar nuclei (karyogamy) to form the nucleus of the central cell was observed. This karyogamy in Agave tequilana occurs before the process of double fertilization (Figure 4c). The central cell nucleus remained located at the same position where both polar nuclei were observed at the chalazal extreme of the embryo sac. The central cell nucleus showed semi-circular to ovate morphologies with an approximate size of 17 µm long and 13 µm wide, and the approximate distance between the nucleus of the egg cell and that of the central cell was 210 µm. After the double fertilization took place at three DAP (to be published elsewhere), the embryo sac started to increase its size and changed its morphology, thus at five DAP, the embryo sac had increased its size to about 280 µm long and 125 µm wide. At the same time, the embryo sac walls near the chalazal end and those that surround the haustorial tube begin to move towards the nucellar tissue (Figure 6a). Early embryo development of Agave tequilana Weber. a) Zygote and early endosperm development. Bar = 10 µm. b) Formation of the chalazal haustorium (curved arrows) Bar = 50 µm. c) Formation of the micropylar haustorium. Bar = 25 µm. e) and f) Immature seed showing the apical and basal cells of the early embryo. Bar = 25 µm. e) and f) Immature seed showing the micropylar haustorium and the chalazal haustorium including the postament. Bar in e = 100 µm and Bar in f = 200 µm. ch = chalaza, m = micropyle, zy = zygote, zyn = zygote, zyn = zygote nucleus, eni = endosperm nucleus, eni = enfile 2: Figure S5). The zygote formation could be observed as the result of the fertilization of the egg cell by one of two sperm nuclei. The zygote formation, the formation of the egg cell by one of two sperm nuclei. disperse cells of the endosperm could be observed (Figure 6a,b, c). The endosperm nucleus forms a cell wall, which generates two cells: a small cell at the chalazal extreme and a large cell at the micropylar extreme that
occupies most of the embryo sac (see Additional file 2: Figure S4). At six DAP, the Agave tequilana zygote enlarges almost 50% maintaining its original width. Meanwhile, the endosperm nuclei continued divisions of the embryo sac. Several nuclear divisions of the endosperm nuclei continued division of the embryo sac. zygote occurred. Likewise, the embryo sac changes in form and size radically, its shape is irregular and resembles that of a heart, the walls surrounding the haustoria: a new one on each side of the chalazal tube still pushing towards the chalaza which in turn included the hypostase which was connected to the former chalazal haustorium (Figure 6b). Similarly, at the micropylar extreme starts the formation of the micropylar haustorium where the zygotic embryo is developed (Figure 6c,d,e). At this stage of development, it was possible to observe well defined chambers, one at the micropylar end and two at the chalazal end (Figure 6e). At eight and nine DAP the first division of the zygote could be observed. This first division gives rise to two cells: one basal cell and one apical cell that would form the embryo proper. Similarly to the zygote nucleus, the embryo proper. that is preserved from the egg cell, resulted in the asymmetric division of the zygote, thus the apical cell was smaller than the basal cell, and possessed a big and well-defined nucleus, while the basal cell seemed to be highly vacuolated (Figure 6d). At this point, the fertilized embryo sac is an immature seed ready toward maturity (Figure 6f). The chalaza-micropyle linear orientation of tetrads in other species of this family has been previously reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1941) and Cave (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1941). However, Wolf (1940) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1941) and Cave (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1941) and Cave (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1941) and Cave (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1941) and Cave (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1941) and Cave (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1941) and Cave (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1941) and Cave (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1941) and Cave (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1948) reported that the formation of tetrads as the most frequent arrangement in Y. aloifolia, while Regen (1948) reported that the formation of tetrads as the most frequent arrangement arrangement arrangement arrangement arrangement arrangement arrangement arrangement arra of meiotic tetrads may take either the "T" or the linear shape in A. virginica and Hesperocallis undulata respectively. In Agave tequilana, only the closest megaspore (FM). This is the case of most angiosperms including Arabidopsis thaliana and Zea mays (Yang et al. 2010) and the close related species Agave virginica (Regen 1941) and Yucca aloifolia (Wolf 1940). However, this megaspore is not always the one that remains viable and is converted to a FM in Y. filamentosa. On the other hand, Piven et al. (2001) observed that the embryo sac of Agave fourcroydes and A. angustifolia develops from the two megaspores closest to the micropylar pole originating an embryo sac of the bisporic Allium-type. The size of the functional megaspore (43 µm long and 25 µm wide) found in the present study was similar to the dimensions reported for Tofieldia glutinosa where the size of the FM was 35-50 µm long and 12-20 µm wide (Holloway and Friedman 2008). The hypostase located at the chalazal pole of the embryo sac in the nucellar tissue is reported as frequent among members of the Asparagaceae (formerly Agavaceae) family (Tilton and Mogensen 1980), and it is probable that it plays an important function in the translocation of nutrients from the ovule to the gametophyte before and after fertilization (Tilton 1980). Furthermore, the three synchronized mitotic divisions without cytokinesis of the functional megaspore were similar to many reported in the literature i.e. in the formation of the maize embryo sac (Huang and Sheridan 1994). The haustorial tube at the chalazal end of the embryo sac observed in Agave tequilana resembles to that reported by Tilton (1978) for Ornithogalum. This haustorial tube may play a role for some kind of a nutritious function and penetrates the nucellar tissue close to the hypostase and the vascular strands, which come from the funiculus (Reed 1903; Watkins 1937; Wolf 1940; Rudall 1997). The cellularized embryo sac consisted of seven cells (eight nuclei): three antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei are found in many one egg cell and two synergids. Occasionally, sacs with less than eight nuclei): three antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the antipodal cells situated at the chalazal end (only their residues could be observed); two polar nuclei close to the ant species and this can be due to a rapid degeneration of the antipodals or they may go unnoticed because they are hidden at the end of the chalazal tube (Maheshwari 1948, 1950). This phenomenon of rapid disintegration of the antipodal cells has been reported for several plant species, such as Agave virginica (Regen 1941), Glycine max (Kennell and Horner 1985), Scilla persica (Svoma and Greilhuber 1988), Triticum aestivum (Zhang et al. 2002), Sargentodoxa cuneata (Wang et al. 2009) and Cichorium intybus (Chehregani et al. 2011). According to Tilton (1978), in angiosperms, antipodals are cells that vary in their behavior in the mature megagametophyte and the only trait they share with each other is their location in the chalazal end of the sac; the antipodals can be ephemeral, degraded shortly after their formation or persist even after fertilization (Williams and Friedman 2004). In Tofieldia glutinosa, antipodals can even proliferate in the maturation stage of the embryo sac, being up to eight antipodal nuclei (Holloway and Friedman 2008). Polar nuclei in Agave tequilana were observed close to the antipodals remains and being similar to that observed in Hemiphylacus alatostylus (Rudall et al. 1997) which are located in the chalazal end within a tube or neck (chalazal haustorium), and in Tofieldia glutinosa where polar nuclei are located in the chalazal end within a tube or neck (chalazal haustorium), and in Tofieldia glutinosa where polar nuclei are located in the chalazal end within a tube or neck (chalazal haustorium), and in Tofieldia glutinosa where polar nuclei are located in the chalazal end within a tube or neck (chalazal haustorium), and in Tofieldia glutinosa where polar nuclei are located in the chalazal haustorium). the micropyle is highly similar to what was observed in the ovular apparatus of Ornithogalum caudatum (Tilton 1978), and the polarization of the nucleus of both synergids towards the micropylar end was similar to what was reported for tobacco (Tian et al. 2005). Furthermore, as in many other angiosperms, the egg cell possesses a highly dense nucleus located towards the chalazal extreme of the embryo sac (Tilton 1978; Mogensen and Suthar 1979; Tian et al. 2005). It was observed that in Agave tequilana, karyogamy of the polar nuclei to form the diploid nucleus of the central cell occurred before the process of double fertilization, similar to that observed in ovules of Capsella
bursa-pastorised towards the central cell occurred before the process of double fertilization, similar to that observed in ovules of Capsella bursa-pastorised towards the central cell occurred before the process of double fertilization, similar to that observed in ovules of Capsella bursa-pastorised towards the central cell occurred before the process of double fertilization, similar to that observed in ovules of Capsella bursa-pastorised towards the central cell occurred before the process of double fertilization, similar to that observed in ovules of Capsella bursa-pastorised towards the central cell occurred before the process of double fertilization, similar to that observed towards the central cell occurred before the process of the central cell occurred before the proces of th (Schulz 1973) and Tofieldia glutinosa (Holloway and Friedman 2008). This diploid nucleus remained at the same place where both polar nuclei were observed in Yucca rupicola (Watkins 1937), Persea americana (Tomer and Gottreich 1976) and Tofieldia glutinosa (Holloway and Friedman 2008) among others. However, Piven et al. (2001) reported for Agave fourcroydes and A. angustifolia that the position of the central cell were located in the center of the embryo sac; or close to the egg apparatus at the micropylar end, a pattern that is present in almost all angiosperms (Tilton 1978; Russell 1993), particularly in Zea mays (Huang and Sheridan 1994) and in Arabidopsis thaliana (Olsen 2004). Double fertilization of the zygote and the endosperm. In this work the typical polarization of the zygote and the endosperm. In this work the typical polarization of the zygote and the endosperm. In this work the typical polarization of the zygote and the endosperm. zygote nucleus resembled those of Capsella bursa-pastoris (Schulz and Jensen 1968), Nicotania tabacum (Mogensen and Suthar 1979) and Arabidopsis thaliana (Mansfield et al. 1991). In this study, the helobial type of endosperm development was similar to the one reported for Hesperocallis undulata (Cave 1948) which has the central cell nucleus at the chalazal extreme of the embryo sac just below of the antipodal cell remains located at the chalazal tube. Maheshwari (1950) reported that when the position of the central cell nucleus is located close to the antipodals, the endosperm type of development will be helobial. At six DAP, the Agave tequilana zygote enlarged almost 50% maintaining its original shape, this enlargement has been previously reported for Arabidopsis thaliana (Bowman et al. 1994) among many others, and the endosperm cells continued dividing before the first division of the zygote. These observations are similar to those in Amaranthus hypochondriacus (Coimbra and Salema 1999), suggesting that the central cell is precocious in regard to its development after fertilization. Around these days, the embryo sac suffers drastic changes in size and with the formation of two chalazal haustoria. There exist several reports about the formation of chalazal haustoria in species belonging to the Asparagales, where the endosperm development invades lateral sections of the proximal nucellus, destroying the lateral tissue and leaving the "postament" at the center of the sac (Rudall 1997). Finally, at nine DAP the first division of the sygote takes place giving rise to one basal cell and one apical cell which is the first cell of the embryo proper. This observed process was similar to what is described for the majority of angiosperms (Lau et al. 2012). Furthermore, the polarity of the embryo sac occurred similarly as in most of flowering plants (Rodríguez-Garay et al. 2000). The Agave teguilana embryo sac development is a monosporic Polygonum-type, showing ephemeral antipodals. In the present work, the detailed embryo sac development, the formation of the zygote, the early embryo formation and the helobial type of endosperm are reported for the first time in this species. knowledge. The results reported here show basic knowledge about the early embryo development and allow new paths for basic and evolutionary studies and breeding programmes where in vitro fertilization, selfing, and intra- and inter-generic hybridization are needed. Agave tequilana presents perfect flowers with six tepals and anthers, an inferior ovary that is divided in three locules, each locule containing two rows of numerous anatropous ovules with axillary placentation (Gentry 1982). The plant material used in this study consisted of inflorescences collected from mature plants growing in the state of Jalisco, which is located in the Tequila appellation of origin in Mexico (DOF 1977), from May to June in the years 2010-2013. At the beginning of the flowering season, panicles per inflorescence or plant). Afterwards, ovules from these young buds were extracted from the ovary using fine forceps and knives under a dissection microscope, and then fixed in a FAA solution (10:5:50:35 formaldehyde: acetic acid: ethanol; distilled water) for 24 hours. After fixation, ovules were transferred to a 70% ethanol solution and stored at 5°C for later staining. In order to maintain cross-pollination to ensure fertilization and embryo formation, the rest of the flower buds. were emasculated before anthesis and removed antheris were kept at room temperature until they matured after one or two days. Mature pollinated from a desiccator at 4°C for future pollinations. Once stigmas were receptive, 10 mature non-pollinated from a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccator at 4°C for future pollen was recovered and stored in a desiccato the ovaries, they were processed following the procedure used for buds. The rest of the flowers with receptive stigmas were hand pollinated using a small paintbrush. Cross-pollination did not assure fertilization. In order to study the embryo sac and zygotic embryo development, 10 immature fruits were collected from panicles at 2, 3, 4, 5, 6, 8 and 9 days after pollination (DAP). Ovules and immature seeds, previously fixed in FAA and kept in 70% ethanol, were stained based on the technique reported by Stelly et al. (1984), in short, a Mayer's-Hematoxylin solution was used for staining during a 10-24-hour period and then treated with a 2% acetic acid solution during a 16-hour period in order to eliminate the excess stain. Ovule samples were washed with a 0.1% sodium bicarbonate solution for 24 hours at room temperature before dehydration. Afterwards, samples were dehydrated in an ethanol series of 25%, 50%, 70%, 85%, 95% and 100% during 15 minutes, and finally, in 100% ethanol during 2 hours. Clarification was carried out through a series of methyl salicylate solution for microscope observation. The samples were analyzed using a Leica DMR microscope (Wetzlar, Germany) coupled to an Evolution OEi camera (Media-Cybernetics, Bethesda, USA), and microphotographs were processed with the Adobe Photoshop Software version CS6 and evenly adjusted for better contrast. We thank V.M. Gómez-Rodríguez and J. M. Rodríguez-Domínguez for microscope technical assistance and I.J. Román-Guzmán, F.J. Cervantes-Hernández and H. Rodríguez-Julián for their assistance with the artwork. We also thank C. Vélez-Gutiérrez (Tequila Cuervo, S.A. de C.V.) for providing Figure 1a. A.G.G.G. is currently a graduate student at Posgrado en Ciencias de la Floricultura, CIATEJ, Guadalajara, Jalisco, México (BEI-AGA-10-8, BEI-AGA-11-8, BEI-AGA-10-8, BEI-AGA-11-8, BEI-A 8). Competing interests The authors declare that they have no competing interests. Authors' contributions AGGG carried out the manuscript. AGM helped with interpretation of data and drafted the study carried out analysis and interpretation of data and drafted the manuscript. All authors read and approved the final manusc Scholar]Angiosperm Phylogeny Group (APG III) An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants. Bot J Linn Soc. 2009;161:105-121. doi: 10.1111/j.1095-8339.2009.00996.x. [CrossRef] [Google Scholar]Bowman JL, Mansfield SG, Koorneef M. Embryogenesis. In: Bowman JL, editor. Arabidopsis an atlas of morphology and development. 1. New York: Springer-Verlag; 1994. pp. 349-401. [Google Scholar]Chasan R, Walbot V. Mechanisms of plant reproduction: questions and approaches. Plant
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